

Web based Test Report of the Interlaboratory Comparison Study on detection of *Salmonella* spp. in samples from the primary production stage (chicken faeces), organised by the EURL-*Salmonella*, study XVII-2014.



TEST REPORT

INTERLABORATORY COMPARISON STUDY ON THE
DETECTION OF SALMONELLA spp. IN SAMPLES FROM THE PRIMARY PRODUCTION
organised by EURL-Salmonella
XVII - 2014

1. Laboratory

Laboratory code	<input type="text"/>
Name contact person	<input type="text"/>
Email address contact person	<input type="text"/>
Name of Laboratory or institute	<input type="text"/>
Country	<input type="text" value="[Select country]"/>

2. Is your laboratory accredited according to ISO 17025

- Yes
 No

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INTERLABORATORY COMPARISON STUDY ON THE
DETECTION OF SALMONELLA spp. IN SAMPLES FROM THE PRIMARY PRODUCTION
organised by EURL-Salmonella
XVII - 2014

1. Laboratory

Laboratory code	<input type="text"/>
Name contact person	<input type="text"/>
Email address contact person	<input type="text"/>
Name of Laboratory or institute	<input type="text"/>
Country	<input type="text" value="[Select country]"/>

2. Is your laboratory accredited according to ISO 17025

- Yes
 No

3. Method

- ISO 6579 (RVS and MKTTn)
 Annex D of ISO 6579 (MSRV)
 Other

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INTERLABORATORY COMPARISON STUDY ON THE
DETECTION OF SALMONELLA spp. IN SAMPLES FROM THE PRIMARY PRODUCTION
organised by EURL-Salmonella
XVII - 2014

1. Laboratory

Laboratory code

Name contact person

Email address contact person

Name of Laboratory or institute

Country

2. Is your laboratory accredited according to ISO 17025

- Yes
 No

3. Planning to become accredited, for the determination of Salmonella?

- Yes
 No

4. Planning of accreditation indicate

Planned year

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4. Date of arrival of the parcels

Date

Time

5. Was your parcel damaged at arrival?

- Yes
 No

6. Start time of storage at +5 °C (chicken faeces)

Date

Time

7. Starting date testing

Date

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4. Date of arrival of the parcels

Date

Time

5. Was your parcel damaged at arrival?

Yes

No

Description of damage

6. Start time of storage at +5 °C (chicken faeces)

Date

Time

7. Starting date testing

Date

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POSITIVE CONTROL SAMPLE

8. What kind of positive control did you use?

- Capsule
- Culti loops
- Culture
- Freeze dried (ampoule)
- Lenticule disc
- Other

9. What was the concentration of the *Salmonella* control sample and what volume of BPW was used for the pre-enrichment ?

CFU/sample

ml BPW used ml

10. Which *Salmonella* serovar did you use in the control sample?

- Enteritidis
- Typhimurium
- Goldcoast
- Nottingham
- Panama
- Poona
- Tranaroa (*Salmonella enterica* subsp. *salamae*)
- Other

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PRE-ENRICHMENT – Buffered Peptone Water (BPW)

11. Medium information BPW


Name manufacturer

Code number manufacturer

pH at the day of use

measured at (°C)


12. Start incubation BPW

Date 

Time

temperature incubator (°C)

13. End incubation BPW

Date 

Time

Temperature incubator (°C)

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SELECTIVE ENRICHMENT - Modified Semi solid Rappaport Vassiliadis medium (MSRV)

14. Medium information MSRV

Name manufacturer	<input type="text"/>
Code number manufacturer	<input type="text"/>
Concentration novobiocin per 1L medium (mg/L)	<input type="text" value="mg/L"/>
pH at the day of use	<input type="text"/>
Measured at (°C)	<input type="text" value="°C"/>

15. Start incubation MSRV

Date	<input type="text"/>
Time	<input type="text" value="hh:mm"/>
temperature incubator (°C)	<input type="text" value="°C"/>

16. End incubation MSRV (24 h)

Date	<input type="text"/>
Time	<input type="text" value="hh:mm"/>
temperature incubator (°C)	<input type="text" value="°C"/>

17. End incubation MSRV (48 h)

Date	<input type="text"/>
Time	<input type="text" value="hh:mm"/>
Temperature incubator (°C)	<input type="text" value="°C"/>

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ISOLATION - Xylose Lysine Desoxycholate medium (XLD)

18. Medium information XLD

Name manufacturer

Code number manufacturer

19. Start of incubation 24 h MSRV/XLD

Date

Time

Temperature incubator (°C)

20. End of incubation 24 h MSRV/XLD

Date

Time

Temperature incubator (°C)

21. Start of incubation 48 h MSRV/XLD

Date

Time

Temperature incubator (°C)

22. End of incubation 48 h MSRV/XLD

Date

Time

Temperature incubator (°C)

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ISOLATION – Second Isolation medium (prescribed)

23. Medium information second isolation medium

Name of the medium


Other

Prescribed incubation temperature in °C

Name manufacturer

Code number manufacturer


24. Start incubation 24 h MSRV/second isolation medium

Date 

Time

Temperature incubator (°C)


25. End of incubation 24 h MSRV/second isolation medium

Date 

Time

Temperature incubator (°C)


26. Start of incubation 48 h MSRV/second isolation medium

Date 

Time

Temperature incubator (°C)

27. End of incubation 48 h MSRV/second isolation medium

Date 

Time

Temperature incubator (°C)

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CONFIRMATION – Nutrient agar

28. Did you streak the colonies on Nutrient agar before starting confirmation?

- Yes
 No

CONFIRMATION of *Salmonella* suspect colonies

29. What media/test did you use for confirmation?

- Biochemical
 Serotyping
 Other

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CONFIRMATION – Nutrient agar

28. Did you streak the colonies on Nutrient agar before starting confirmation?

- Yes
 No

CONFIRMATION of *Salmonella* suspect colonies

29. What media/test did you use for confirmation?

- Biochemical
 Serotyping
 Other

30. Biochemical

- Triple sugar/iron agar (TSI)
 Urea Agar (UA)
 L-Lysine decarboxylation medium (LDC)
 Galactosidase
 Voges-Proskauer (VP)
 Indole

Identification kit (name of the kit)

Other

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CONFIRMATION – Nutrient agar

28. Did you streak the colonies on Nutrient agar before starting confirmation?

- Yes
 No

CONFIRMATION of *Salmonella* suspect colonies

29. What media/test did you use for confirmation?

- Biochemical
 Serotyping
 Other

30. Serotyping

- O antigens
 H antigens
 Vi antigens
 Other

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DETECTION BY PCR (optional)

31. Did you use PCR in this study

- Yes
 No

32. What kind of PCR?

- Real time PCR
 Other PCR

33. Is the PCR used commercially available?

- Yes
 No

34. Is the PCR validated ?

- Yes
 No

35. Is the PCR published in the open literature ?

Reference literature

36. Do you use the PCR routinely ?

- Yes
 No

37. At what moment did you start with the extraction/detection?

- Before pre-enrichment in BPW
 After pre-enrichment in BPW
 After selective enrichment on MSRV
 Other

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DETECTION BY PCR (optional)

31. Did you use PCR in this study

- Yes
 No

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DETECTION BY PCR (optional)

31. Did you use PCR in this study

- Yes
 No

32. What kind of PCR?

- Real time PCR
 Other PCR

33. Is the PCR used commercially available?

- Yes
 No

34. Name of PCR, manufacturer and batch used in the study

Manufacturer

Batch

35. Is the PCR validated ?

- Yes
 No

36. For which matrix/matrices and by which organisation?

Matrices

Validated by

Ref. number

37. Is the PCR published in the open literature ?

Reference literature

38. Do you use the PCR routinely ?

- Yes
 No

39. How many samples did you test for Salmonella using this PCR in 2013 ?

Number

40. At what moment did you start with the extraction/detection?

- Before pre-enrichment in BPW
 After pre-enrichment in BPW
 After selective enrichment on MSRV
 Other

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Table 1 Confirmed results for Salmonella after selective enrichment on MSRV (samples B1-B18 & C1-C3)

Indicate:

- 1 for positive confirmed result for Salmonella
- 0 for negative (confirmed) result for Salmonella

39. Table 1

	24 h MSRV/XLD	24 h MSRV/Second isolation medium	48 h MSRV/XLD	48 h MSRV/Second isolation medium
B1	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B2	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B3	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B4	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B5	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B6	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B7	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B8	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B9	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B10	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B11	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B12	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B13	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B14	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B15	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B16	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B17	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
B18	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
C1	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
C2	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾
C3	[Select result] ▾	[Select result] ▾	[Select result] ▾	[Select result] ▾

Table 2 Results of Salmonella detection using PCR (samples B1-B18 & C1-C3)

Indicate:

1 for positive PCR result for Salmonella
0 for negative PCR result for Salmonella

40. Table 2

	PCR
B1	[Select result] ▼
B2	[Select result] ▼
B3	[Select result] ▼
B4	[Select result] ▼
B5	[Select result] ▼
B6	[Select result] ▼
B7	[Select result] ▼
B8	[Select result] ▼
B9	[Select result] ▼
B10	[Select result] ▼
B11	[Select result] ▼
B12	[Select result] ▼
B13	[Select result] ▼
B14	[Select result] ▼
B15	[Select result] ▼
B16	[Select result] ▼
B17	[Select result] ▼
B18	[Select result] ▼
C1	[Select result] ▼
C2	[Select result] ▼
C3	[Select result] ▼

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41. Remarks and Comment(s)

42. Name of person (s) carrying out the Comparison study

Name

43. Name of the person in charge

Name

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Review and submit

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